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Periodontitis is associated with the increased levels of visfatin: a meta-analysis



Junfei Zhu^{1†}, Suhan Zhang^{2†}, Jing Shi³, Ning ning³, Ying Wei^{4*†} and Ye Zhang^{1*†}

Abstract

Objective Periodontitis is a common inflammatory disease associated with systemic factors. Visfatin is a pleiotropic adipokine that exerts metabolic and immune functions. Studies have shown visfatin played roles in the development of periodontitis. The present study aims to compare the levels of visfatin in body fluids including serum, saliva, and gingival crevicular fluid (GCF) between periodontitis patients and healthy individuals, and to elucidate the alteration of visfatin levels after periodontal treatments.

Materials and methods The database searched included Pubmed, Embase, Web of Science, and Cochrane Library. According to the Eligibility criteria, the records were screened and the eligible studies were included. The methodological qualities of the included case-controlled studies were assessed according to the Newcastle–Ottawa scale (NOS). The Methodological Index for Nonrandomized Studies (MINORS) was applied for assessing the qualities of the included clinical trials. The statistical analyses were processed using STATA 15.0.

Results Twenty-three studies were included in the statistical analyses. The meta-analysis showed significantly elevated visfatin levels of GCF, serum, and saliva in the periodontitis population compared with the controls (GCF: SMD = 5.201, 95% CI: 3.886-6.516, Z = 7.75, P < 0.05; Serum: SMD = 7.417, 95% CI: 3.068-11.767, Z = 3.34, P = P < 0.05; Saliva: SMD = 2.683, 95% CI: 1.202-4.163, Z = 3.34, P < 0.05). Visfatin levels of saliva serum and GCF were significantly decreased after periodontal treatment.

(Saliva: SMD = -1.338, 95% CI: -2.289—0.487, Z = 39.77, P < 0.05; Serum: SMD = -2.890, 95% CI: -5.300-0.480, Z = 2.35, P < 0.05; GCF: SMD = -6.075, 95% CI: -11.032—1.117, Z = 2.40, P = 0.016; I 2 = 95.9%, P < 0.05).

Conclusions Periodontitis elevated the visfatin levels in GCF, serum, and saliva. Additionally, GCF, serum, and saliva visfatin levels could be reduced after periodontal treatment.

Keywords Visfatin, Periodontitis, Periodontal treatment, Gingival crevicular fluid, Meta-analysis

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Introduction

Periodontitis is a common inflammatory disease induced by polymicrobial factors. The consequences of periodontal disease are frequently presented as gingival infection, alveolar bone destruction, as well as periodontal attachment loss [1]. The status of periodontitis strongly affected the quality of life. The destructed periodontal tissues not only give an annoying smell but also leads to aesthetic compromise, which influenced social life [2]. Additionally, the masticatory function declined, which brought difficulties in nutrition intake. Moreover, periodontitis has been reported to be associated with multiple systemic diseases, such as diabetes, cardiovascular disease, and cancer [3, 4]. The etiology of periodontitis has been extensively studied. Although the inflammation of periodontal tissue is supposed to be initialed by bacteria, the disproportionate and unbalanced host response to the microbial film and the inability of the host to resolve the inflammatory process is considered to be responsible for the occurred periodontal damage [5].

Adipokines are a complex cohort of cytokines produced by adipocytes, including leptin, adiponectin, resistin, and visfatin. etc. [6] It has been widely demonstrated that adipokines are involved in multiple internal events, such as energy metabolism, wound healing, as well as inflammatory reactions [7]. Visfatin, also identified as the pre-Bcell colony-enhancing factor (PBEF), is a 52-kDa protein promoting pre–B cell colony release from lymphocytes and improving the maturation of B lymphocytes [8]. As a pleiotropic cytokine, visfatin has roles in many processes, including insulin mechanisms, apoptosis, and inflammation regulation [9]. The association between visfatin and the pathogenesis of several systemic diseases, such as type 2 diabetes mellitus, polycystic ovary syndrome, and cardiovascular diseases has also been reported [10]. Additionally, in the process of inflammation, visfatin induces pro-inflammatory cytokines including tumor necrosis factor-alpha (TNF- α), interleukin (IL)- 1,6, and 8 [11], which played a critical role in the initiation and development of systemic inflammation from neutrophils, lymphocytes, and macrophages.

The relationship between adipokines and periodontitis has been reported. Previously, the present author published a meta-analysis regarding the association between periodontitis and serum levels of leptin and adiponectin, finding that elevated serum leptin and decreased serum adiponectin were related to periodontitis [12]. Recent year studies have shown that visfatin also played roles in the development of periodontitis. Multiple studies have been published to discover the possible links between visfatin and periodontitis, However, an evidence-based summative study was needed to provide a more precise evaluation. The present study gathered individual study results into a quantitative estimation, the aim was to compare the levels of visfatin in body fluids including serum, saliva, and gingival crevicular fluid (GCF) between periodontitis patients and healthy individuals, and to elucidate the alteration of visfatin levels after periodontal treatments.

Methods

We conducted the present meta-analysis following the Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement [13]. The protocol of the present study was registered in the International platform of registered systematic review and meta-analysis protocols (INPLASY), the registration number is INPLASY202380125.

Literature search

The database searched included Pubmed, Embase, Web of Science, and Cochrane Library, up to December 2022. The details of search strategies are presented in Table 1.

Eligibility criteria

The inclusion criteria for the included studies were as follows: (1) the observational studies comparing visfatin levels in serum, saliva, or GCF between periodontitis patients and periodontally healthy individuals; (2) Clinical trials comparing visfatin levels in serum, saliva, or GCF before and after periodontal treatments; (3) Studies

 Table 1
 The details of search strategies

Pubmed	("periodontal"[Title/Abstract] OR "periodontitis"[Title/Abstract]) AND ("visfatin"[Title/Abstract] OR "nicotinamide phosphoribosyltran sferase"[Title/Abstract] OR "NAMPT"[Title/Abstract] OR "pre b cell colony enhancing factor"[Title/Abstract] OR "PBEF"[Title/Abstract])
Embase	(visfatin:ab,ti OR 'nicotinamide phosphoribosyltransferase':ab,ti OR nampt:ab,ti OR 'pre b cell colony enhancing factor':ab,ti OR pbef:ab,ti) AND (periodontitis:ab,ti OR periodontal:ab,ti)
Web of science	(TS = (periodontal) OR TS = (periodontitis)) AND (TS = (Visfatin) OR TS = (nicotinamide phosphoribosyltransferase) OR TS = (NAMPT) OR TS = (pre b cell colony enhancing factor) OR TS = (PBEF))
Cochrane Library	((periodontal):ti,ab,kw OR (periodontal disease):ti,ab,kw (Word variations have been searched))AND ((visfatin):ti,ab,kw OR (nicotina- mide phosphoribosyltransferase):ti,ab,kw OR (NAMPT):ti,ab,kw OR (pre b cell colony enhancing factor):ti,ab,kw OR (PBEF):ti,ab,kw (Word variations have been searched))

with sufficient data for the statistical analyses; and (4) Studies in English or Chinese.

The exclusion criteria were as follows: (1) Conference abstracts; (2) Trial registry records; and (3) those with repeated data.

Records screen

Search results were downloaded to EndNote. According to the Eligibility criteria, the records were collated and separately reviewed by two independent authors. A third author was consulted when consensus on appropriateness was debated. The titles and abstracts were screened first and the full-text paper screen was conducted next.

Data extraction

The information on study characteristics and outcomes was extracted, including (1) Name of the first author and year of publication; (2)Country; (3) study design; (4) Group and size; (5)Age; (6) Body mass index (BMI); (7) Systemic conditions; (8)Type of periodontal treatments; (9) The time interval after periodontal treatment; and (10) Serum, saliva, or GCF levels of visfatin.

Quality assessment

The methodological qualities of the included case-controlled studies were assessed according to the Newcastle-Ottawa scale (NOS). Three dimensions including selection, comparability, and exposure were considered. In the section on comparability, the most important factor was identified to be BMI. Eight items were graded, with a score range of 0-9 points. Overall, studies with final scores of 1-3, 4-6, and 7-9 were considered to be of low, moderate, and high gualities [14]. Additionally, the Methodological Index for Nonrandomized Studies (MINORS) was applied for assessing the qualities of the included clinical trials. A total of 12 items were considered, and 0-2 points could be graded for each item. As in the present study we only evaluated the quality of singlearm designs, the first 8 items were employed. The final scores ranged from 0 to 16 points. Studies with scores 0-5, 6-10, and 11-16 were considered to be of low, moderate, and high qualities [12].

Meta-analysis

The data of visfatin levels were presented as mean $(M) \pm$ standard deviation (SD). The results in the form of median (minimum – maximum) were shifted to $M \pm$ SD according to the estimation method reported by Hozo et al. [15]. The standard mean difference (SMD) and corresponding 95% confidence interval (CI) were calculated. Heterogeneity was estimated by chi-square and I². Random-effects model was used when the P value < 0.05 and I² > 50%. Sensitivity analyses were performed to test the

robustness. The statistical analyses were processed using STATA 15.0.

Results

Characteristics and quality assessment

The flow diagram of the selection process was presented in Fig. 1. Finally, 23 studies were gained according to the eligibility criteria, including 14 case-controlled studies, 3 clinical trials, and 6 studies that employed both clinical trials and case-controlled designs. The origin of the studies included Iran, Saudi Arabia, Turkey, China, India, Iraq, and the USA. The descriptive information characteristic was presented in Table 2. 20 case-controlled studies were evaluated through NOS. The final scores ranged from 4 to 8 (Table 3). 17 studies were of high quality. MINORS was employed for the 9 clinical trials in the present study. final scores ranged from 11 to 15, indicating all of the 9 clinical trials were of high quality (Table 4).

Meta-analysis

Eleven studies reported the comparison of visfatin levels in GCF between periodontitis patients and healthy individuals. The meta-analysis showed significantly increased GCF visfatin levels in the periodontitis population (SMD=5.201, 95% CI: 3.886-6.516, Z=7.75, P < 0.05; I²=93.0%, P < 0.05 Fig. 2A). 5 studies reported the comparison of visfatin levels in serum between periodontitis patients and healthy individuals. Also, the meta-analysis showed significantly increased serum visfatin levels in the periodontitis population (SMD = 7.417, 95% CI: 3.068–11.767, Z=3.34, P=0.001; I²=97.7%, P< 0.05 Fig. 2B). We found the study conducted by Chen, F et al. [19] contributed to the heterogeneity. When the study was removed, the heterogeneity was significantly decreased, and the result was not influenced (SMD = 8.005, 95% CI: 7.058-8.953, Z = 16.56, P < 0.05; I 2 =54.4%, P=0.087). 5 studies reported the comparison of visfatin levels in saliva between periodontitis patients and healthy individuals. The meta-analysis showed significantly increased saliva visfatin levels in the periodontitis population (SMD=2.683, 95% CI: 1.202-4.163, Z=3.34, P = 0.001; I² = 93.3%, P < 0.05 Fig. 2C).

Six clinical trials reported the change in saliva visfatin levels after periodontal treatments. The results of the meta-analysis showed a significantly declined saliva visfatin levels after periodontal treatment compared with the baseline (SMD=-1.338, 95% CI: -2.289— 0.487, Z=39.77, P=0.003; I ²=87.4%, P<0.05 Fig. 3A). the study conducted by Saseendran, G et al. [34] contributed to the heterogeneity. When the study was removed, the heterogeneity was significantly decreased, and the result was not influenced (SMD=-0.710, 95% CI:



Fig. 1 The flow diagram of the selection process

-1.023—0.397, Z=16.56, P < 0.05; I²=17.8%, P=0.301). 3 studies for each described the change of visfatin levels in serum and GCF after periodontal treatment. The results of the meta-analysis also showed significantly declined serum and GCF visfatin levels after periodontal treatment compared with the baseline (Serum: SMD=-2.890, 95% CI: -5.300-0.480, Z=2.35, P=0.0019; I 2=95.4%, P < 0.05 Fig. 3B; GCF: SMD=-6.075, 95% CI: -11.032—1.117, Z=2.40, P=0.016; I 2=95.9%, P < 0.05 Fig. 3C).

The sensitivity analyses were proceeded by removing a single study each time and observing the influence of each study on the result. The results of sensitivity analyses were presented in Fig. 4, according to which the robustness of the results was confirmed.

Publication bias

Egger's linear regression test was used to examine the potential publication bias. We found a significant publication bias in the comparison of visfatin levels in GCF between periodontitis patients and healthy individuals (P < 0.05). For other comparisons, Egger's linear

regression test was not conducted because the number of included studies was limited (n < 10).

Discussion

Increasing evidence in the literature indicated the associations between adipocytokines and periodontitis. In 2016, Zohaib Akram et al. published the first metaanalysis researching the cytokine profile in periodontitis patients with and without Obesity, finding increased resistin levels in obese individuals with periodontitis than nonobese periodontitis subjects [39]. Another evidencebased study conducted by the team found that periodontitis patients were presented with elevated levels of GCF or serum resistin, indicating resistin might be used as a potential biomarker for chronic periodontitis [40]. In 2017, the author of the present study published a systematic review and meta-analysis regarding the circulating leptin and adiponectin levels in the periodontitis population, the results showed elevated serum levels of leptin and decreased serum levels of adiponectin were found in

Author	Year	Country	Study design	Group and size	Age	BMI	Systematic conditions	Periodontal treatment	Time interval	Sample
Abolfazli, N [16]	2015	Iran	Single-armed clinical trial	Periodontitis (18)	41.16±7.59	24.35 ± 0.63	Healthy	SRP	4 weeks	Serum and saliva
Bahammam, M. A [17]	2018	Saudi Arabia	Case-controlled	Control(20)/ Periodontitis and T2DM (20)/ Periodontitis(20)/ Periodontitis and T2DM using simvastatin(20)	32–50	¢ Z	/MbCT/ydtaeH MbCT/ydtaeH	Ч Ч	Ч	GCF
Çetiner, D [18]	2019	Turkey	Case-controlled	Periodontitis (9)/Control(10)/ Obese and peri- odontitis (21)/ Obese control (10)	44.67±10.87/46.50±1.2.0 (Non-obese); 44.22±6.7/27.80±3.12(Obese)	BMI < 25 (Non-obese); BMI ≥ 30(Obese)	Healthy/Healthy/ Obese/Obese	۲ ۲	۲ Z	GCF
Chen, F. [19]	2018	China	Case-controlled	Periodontitis (79)/Control (25)	Male (Female): 30(27) /30(25)	NA	Healthy/Healthy	NA	NA	Serum
Coutinho, A [20]	2021	India	Case-controlled	Periodontitis (20)/Control (20)	20-50	18.5–29.9	Healthy/Healthy	NA	NA	Saliva
Hamdi, A. Q [21]	2021	Iraq	Case-controlled	Periodontitis and T2DM (41)/ Control (49)	60±14/61.8±12.5	NA	T2DM/Healthy	AN	AN	Serum
Mishra, V [22]	2016	India	Case-controlled; Clinical trial	Control(14)/Peri- odontitis (14)/ Periodontitis and T2DM (14)	32.43/41.7/48.07	NA	Healthy/Healthy/ T2DM	SRP	4 weeks	GCF
Mohamed, H. G [23]	2015	USA	Case-controlled	T2DM and periodon- titis (54)/T2DM control(24)/ Periodontitis (30) Control (44)	54.76±1.38/50.79±2.09/55.37±1.7 7/47.15±1.56	٩	Healthy/Healthy/ T2DM/T2DM	Ч И	٩	GCF
Mopidevi, A [24]	2019	India	Case-controlled; Single-armed clinical trial	Periodontitis (10)/Control (10)	43.9±9.65/44.1±8.41	24.68±2.65/24.37±2.54	Healthy/Healthy	SRP and Surgi- cal periodon- tal therapy	12 weeks	Saliva
Özcan, E.(A) [25]	2016	Turkey	Case-controlled; Single-armed clinical trial	Periodontitis (17)/Control (15)	41.52±6.34 /44.06±5.36	24.12 ± 0.78/ 23.62 ± 1.53	Healthy/Healthy	SRP	3 months and 6 months	Saliva
Özcan, E.(B) [26]	2016	Turkey	Case-controlled	Periodontitis (27)/Control (18)	41.59 ± 7.23/37.72 ± 9.02	24.74 ± 1.22/24 ± 1.04	Healthy/Healthy	NA	NA	GCF
Özcan, E [<mark>27</mark>]	2015	Turkey	Case-controlled	Periodontitis (25)/Control (23)	32.56 ± 7.92/34.50 ± 7.09	23.50 (21.20– 26.15)/24.55(21.92–28.17)	Healthy/Healthy	NA	NA	Saliva
Paul, R [28]	2020	India	Case-controlled	Periodontitis (30)/Control (30)	30-55	NA	Healthy/Healthy	NA	NA	GCF

 Table 2
 Characteristics of the included studies

Table 2 (con	tinued)	_								
Author	Year	Country	Study design	Group and size	Age	BMI	Systematic conditions	Periodontal treatment	Time interval	Sample
Pradeep, A. R [29]	2011	India	Case-controlled	Periodontitis (15)/Control (10)/ Gingivitis(15)	32.6±7.5/28.70±3.6/28.5±3.1	NA	Healthy/Healthy	NA	NA	Serum and GCF
Pradeep, A. R [30]	2012	India	Case-controlled	Control(10/ Periodontitis and T2DM(10)/ Periodontitis (10)	31.33 ± 3.478/38.80 ± 5.634/32.6 0 ± 7.59	27.28±2.41/28.46±2.72 /28.59±3.14	Healthy/T2DM/ Healthy	ЧN	ЧN	Serum and GCF
Raghavendra, N. M [31]	2012	India	Case-controlled; Single-armed clinical trial	Periodontitis (15)/Control (15)	38.80 ± 5.634/31.33 ± 3.478	NA	Healthy/Healthy	SRP	8 weeks	Serum and GCF
Rezaei, M [32]	2019	Iran	Case-controlled	SLE and peri- odontitis (15)/ SLE control(15)/ Periodontitis (15) Control (15)	44.17±3.21/44.89±3.07/45.03±3.1 4/44.99±3.26	21.89±1.83/22.02±1.17/23. 05±1.65/22.65±2.10	SLE/SLE/Healthy/ Healthy	AN	AN	GCF
Saljoughi, F [33]	2020	Iran	Case-controlled	PCOS and periodon- titis (30)/PCOS control(25)/ Periodontitis (23) Control (32)	45.2±3.2/45.3±3.0/45.3±3.1/45 .5±3.3	22.43±1.79/22.13±1.58/22. 38±1.71/22.07±1.50	PCOS/PCOS/ Healthy/Healthy	NA	٩	GG
Saseendran, G [34]	2021	India	Case-controlled; Clinical trial	Control (16)/ Gingivitis(16)/ Periodontitis(16)	25–50	NA	Healthy/Healthy/ Healthy	SRP	2 months	Saliva
Tabari, Z. A [35]	2014	Iran	Case-controlled	Periodontitis (20)/Control (20)	$38.45 \pm 9.98/33.85 \pm 6.84$	23.66±2.99/21.88±2.56	Healthy/Healthy	٨٨	NA	Saliva
Tabari, Z. A [36]	2015	Iran	Case-controlled; Single-armed clinical trial	Periodontitis (20)/Control (20)	38.45±9.98/33.85±6.84	23.66±2.99/21.88±2.56	Healthy/Healthy	SRP	1 months	Saliva
Wu,Y [37]	2015	China	Randomized clinical trial	T2DM and peri- odontitis (23)/ T2DM and peri- odontitis (23)	54.09±6.57/55.52±5.22	22.22±0.64/22.14±0.72	T2DM/T2DM	SRP	3 months and 6 months	Serum and GCF
Ziaei, N [38]	2020	Iran	Single-armed clinical trial	T2DM and peri- odontitis (20)	51.85±6.42	26.8±1.5	T2DM	SRP	3 months	Saliva

 Table 3
 Quality assessment for case-controlled studies

Author and year	Selection	Comparability	Exposure	Total
Bahammam, M. A 2018 [17]	••0•	0	••0	7/9
Çetiner, D. 2019 [18]	$\bullet \bullet \circ \bullet$	••		8/9
Chen, F. 2018 [19]	$\bullet \circ \bullet \bullet$	$\bigcirc igodot$		7/9
Coutinho, A 2021 [20]	●00●	••	$\bullet \bullet \circ$	6/9
Hamdi, A. Q. 2021 [21]	●00●	00	$\bullet \bullet \circ$	4/9
Mishra, V. 2016 [22]	●00●	$\bigcirc igodot$		5/9
Mohamed, H. G. 2015 [23]	●00●	$\bigcirc igodot$		5/9
Mopidevi, A. 2019 [24]	●00●	••		6/9
Özcan, E (A). 2016 [25]	●00●	••	$\bullet \bullet \circ$	6/9
Özcan, E.(B). 2016 [26]	●00●	••		6/9
Özcan, E. 2015 [27]	●00●	••		6/9
Paul, R 2020 [<mark>28</mark>]	●00●	$\bigcirc igodot$		5/9
Pradeep, A. R. 2011 [29]	●00●	$\bigcirc igodot$		5/9
Pradeep, A. R. 2012 [30]	●00●	••		6/9
Raghavendra, N. M. 2012 [31]	●00●	$\bigcirc igodot$		5/9
Rezaei, M. 2019 [32]	●00●	••		6/9
Saljoughi, F. 2020 [<mark>33</mark>]	●00●	••		6/9
Saseendran, G. 2021 [34]	●00●	$\bigcirc igodot$		5/9
Tabari, Z. A. 2014 [35]	●00●	••		6/9
Tabari, Z. A. 2015 [<mark>36</mark>]	●00●	••		6/9

\bullet = the item was fulfilled; \bigcirc = the item was not fulfilled

periodontitis patients, and periodontal treatment influenced the circulating leptin and adiponectin [12].

The present study is the first meta-analysis concentrated on the association between visfatin and periodontitis. 23 studies were examined in the present study, and the results showed that visfatin levels in GCF, serum, and saliva were significantly elevated in the periodontitis population. Additionally, we also found that GCF, serum, and saliva visfatin levels could be reduced after periodontal treatment.

Human visfatin was first discovered in 1994 as a novel gene isolated from a human peripheral blood lymphocyte cDNA library [41]. Visfatin has been reported as the pleiotropic mediator that exerts metabolic and immune functions [42]. Subsequent work has shown visfatin to be of potent destructive and proinflammatory properties. The protein played a key role in the persistence of inflammation through the inhibition of apoptosis and neutrophils [43]. A variety of inflammatory processes and systematic disorders have been associated with the upregulated expression of visfatin, such as sepsis [44], inflammatory bowel diseases [45], rheumatoid arthritis [46], diabetes [47], as well as cardiovascular diseases [48].

Periodontitis is a common bacterial infection associated with multiple risk factors. Despite the microorganisms' roles as etiologic agents, systemic conditions and inflammatory situations have also contributed to the prevalence and severity of periodontitis [49]. Periodontal supporting tissues were destroyed in periodontitis through the main periodontal pathogens Porphyromonas gingivalis (Pg), Fusobacterium nucleatum (Fn), and the toxic products. On the other hand, the systemic imbalance of pro-inflammatory/anti-inflammatory immune regulation also plays an important role in bone tissue destruction. Studies have shown that visfatin can affect bone metabolism and is a risk factor for osteoporosis [50]. In the present study, we found elevated visfatin levels in body fluids in periodontitis patients, indicating that visfatin might be involved in the inflammatory responses of periodontitis.

Among the 3 kinds of physiological fluids investigated in the present study, GCF was mostly reported. GCF is the fluid and the inflammatory exudate secreted from the blood vessels in the gingival corium, subjacent to the epithelium lining of the dentogingival space. Cytokines presented in GCF have been proposed as potential diagnostic or prognostic markers of periodontal destruction [51]. The present study found elevated visfatin in GCF in periodontitis. The increased visfatin in CGF could be secreted by the inflamed periodontal ligament cells or the activated Porphyromonas gingivalis [28]. In sequence, the

Author and year	1	2	3	4	5	6	7	8	Total
Abolfazli, N 2015 [16]	•	O	•	•	•	O	•	0	12/16
Mishra, V. 2016 [22]	•	O	•	•	•	O	•	•	14/16
Mopidevi, A. 2019 [24]	•	0	•	•	•	•	•	O	13/16
Özcan, E.(A) 2016 [25]	•	O	•	•	•	•	•	0	13/16
Raghavendra, N. M. 2012 [31]	•	0	•	•	•	O	•	0	11/16
Saseendran, G2021 [34]	•	O	•	•	•	O	•	0	14/16
Tabari, Z. A. 2015 [<mark>36</mark>]	•	O	•	•	•	O	•	0	14/16
Wu, Y. 2015 [37]	•	O	•	•	•	•	•	0	15/16
Ziaei, N. 2020 [<mark>38</mark>]	•	0	•	•	•	•	•	0	14/16

 Table 4
 Quality assessment for clinical trials

 \bullet = the item was reported and adequate; \bigcirc = the item was not reported; \bullet = the item was reported but inadequate



Fig. 2 GCF, serum, and saliva visfatin levels between periodontitis patients and healthy individuals. A GCF; B Serum; C Saliva



Fig. 3 The change of visfatin levels after periodontal treatment. A Saliva; B Serum; C GCF



Fig. 4 Sensitive analysis. A GCF visfatin; B Serum visfatin; C Saliva visfatin; D The change of saliva visfatin after periodontal treatment; E The change of serum visfatin after periodontal treatment; F The change of GCF visfatin after periodontal treatment

locally secreted high-level visfatin entered the peripheral blood circulation through the epithelium of the periodontal pocket, the proliferating capillaries, as well as the loose connective tissue. In comparison to GCF, saliva can be collected in an easy, non-invasive, time-saving manner with minimally trained personnel [52]. Salivary proteomics has been a growing topic of research due to its characteristic to be a diagnostic tool for both oral and systemic diseases. And GCF is considered to be a better source for assaying specific dental or periodontal biomarkers of oral conditions due to its ability to be site specific [53]. Additionally, the present study also suggested the upregulated serum visfatin levels in the population with periodontitis, indicating that periodontal inflammation may modulate the systemic levels of visfatin. On the one hand, the increased circulating visfatin could be induced by the locally secreted high-leveled GCF and saliva visfatin. Additionally, another possible explanation for the finding is that peripheral B cells, T cells, monocytes, macrophages, and neutrophils, rather than adipocytes, release visfatin in the body because systematic inflammation could be activated by periodontitis [54].

The limitations of the presented study include the lack of enough included studies, especially randomized controlled studies. Although 9 clinical trials were included, only 3–6 studies were evaluated in each comparison for the change of visfatin levels after periodontal treatment. And, only single-armed results were applied in the meta-analysis to ensure consistency. Besides, significant heterogeneities were observed, and in some comparisons, we failed to find the exact source of heterogeneity, therefore the results should be interpreted with caution. Moreover, although the present study o included studies with case-controlled designs, the results of each study were considered cross-sectional, therefore the cause-effect relationship between visfatin and periodontitis could not be perfectly elucidated.

In conclusion, the present study supports the findings that periodontitis elevated the visfatin levels in GCF, serum, and saliva. Additionally, GCF, serum, and saliva visfatin levels could be reduced after periodontal treatment. Further large-scale homogeneous studies were still needed to prove if visfatin could be used as a potential biomarker to predict periodontitis.

Acknowledgements

Not applicable.

Authors' contributions

Junfei Zhu and Suhan Zhang contributed to the conception of the study; Junfei Zhu performed the meta-analysis and Suhan Zhang wrote the manuscript. Ning Ning and Jing Shi contributed to the records screen and data extraction; Ying Wei and Ye Zhang helped to check and revise the manuscript.

Funding

No funding was obtained for this study.

Availability of data and materials

The datasets used and/or analyzed during the current study available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 24 May 2023 Accepted: 4 September 2023 Published online: 26 October 2023

References

- Slots J. Periodontitis: facts, fallacies and the future. Periodontol 2000. Periodontol. 2017;75(1):7–23.
- 2. Wang Z, et al. The role of adiponectin in periodontitis: Current state and future prospects. Biomed Pharmacother. 2021;137:111358.
- Nwizu N, Wactawski-Wende J, Genco RJ. Periodontal disease and cancer: Epidemiologic studies and possible mechanisms. Periodontol 2000. 2020;83(1):213–33.
- Sanz M, et al. Periodontitis and cardiovascular diseases: Consensus report. J Clin Periodontol. 2020;47(3):268–88.
- Baeza M, et al. Effect of periodontal treatment in patients with periodontitis and diabetes: systematic review and meta-analysis. J Appl Oral Sci. 2020;28:e20190248.
- Giardullo L, et al. Adipokine role in physiopathology of inflammatory and degenerative musculoskeletal diseases. Int J Immunopathol Pharmacol. 2021;35:20587384211015030.
- Unamuno X, et al. Adipokine dysregulation and adipose tissue inflammation in human obesity. Eur J Clin Invest. 2018;48(9):e12997.
- Zhang Y, et al. Visfatin is regulated by interleukin-6 and affected by the PPAR-γ pathway in BeWo cells. Mol Med Rep. 2019;19(1):400–6.
- Cheng Q, et al. Visfatin inhibits apoptosis of pancreatic β-cell line, MIN6, via the mitogen-activated protein kinase/phosphoinositide 3-kinase pathway. J Mol Endocrinol. 2011;47(1):13–21.
- Bayani M, Pourali M, Keivan M. Possible interaction between visfatin, periodontal infection, and other systemic diseases: A brief review of literature. Eur J Dent. 2017;11(3):407–10.
- 11. Kim SR, et al. Upregulation of thromboxane synthase mediates visfatininduced interleukin-8 expression and angiogenic activity in endothelial cells. Biochem Biophys Res Commun. 2012;418(4):662–8.
- Zhu J, et al. Association of circulating leptin and adiponectin with periodontitis: a systematic review and meta-analysis. BMC Oral Health. 2017;17(1):104.
- Moher D, et al. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PLoS Med. 2009;6(7):e1000097.
- Zhu J, et al. Interleukin-6-174G/C Polymorphism Contributes to Periodontitis Susceptibility: An Updated Meta-Analysis of 21 Case-Control Studies. Dis Markers. 2016;2016:9612421.
- Hozo SP, Djulbegovic B, Hozo I. Estimating the mean and variance from the median, range, and the size of a sample. BMC Med Res Methodol. 2005;5:13.
- Abolfazli N, Jabali S, Saber FS, Babaloo Z, Shirmohammadi A. Effect of non-surgical periodontal therapy on serum and salivary concentrations of Visfatin in patients with chronic periodontitis. J Dent Res Dent Clin Dent Prospect. 2015;9(1):11–7. https://doi.org/10.15171/jodd.2015.003.
- Bahammam MA, Attia MS. Effects of systemic simvastatin on the concentrations of Visfatin, tumor necrosis factor-a, and interleukin-6 in gingival crevicular fluid in patients with type 2 diabetes and chronic periodontitis. J Immunol Res. 2018;8481735. https://doi.org/10.1155/2018/8481735.
- Çetiner D, Uraz A, Öztoprak S, Akça G. The role of visfatin levels in gingival crevicular fluid as a potential biomarker in the relationship between obesity and periodontal disease. J Appl Oral Sci. 2019;27:e20180365. https://doi.org/10.1590/1678-7757-2018-0365.
- Chen F, et al. Detection of levels of serum visfatin and PGE2 in patients with periodontitis and their relationships with activity of periodontitis. J Jilin Univ Med Ed. 2018;44(3):563–7.
- Coutinho A, Reddy N, Chatterjee A, Khan MI. The Role of Visfatin (Adipocytokine) Biomarker in Oral Health and Diseases among Nonobese Indian Population: A Proteomic Assay. 2021;8(3):104–8. https://doi.org/10. 1055/s-0041-1728690.
- Hamdi AQ, Sarhat ER, Ali NH, Sarhat TR. Evaluation of Lipocalin-2 and Visfatin, and Vitamin (D,C, and E) in Serum of Diabetic Patients with Chronic Periodontitis. Indian J Forensic Med Toxicol. 2021;15(2):1668–74. https:// doi.org/10.37506/ijfmt.v15i2.14578.
- 22. Mishra V, Shettar L, Bajaj M, Math AS, Thakur SL. Interlinking Periodontitis and Type 2 Diabetes Mellitus by Assessment of Crevicular Visfatin Levels

in Health and in Disease Before and After Initial Periodontal Therapy. J Clin Diagn Res. 2016;10(8):ZC67–71. https://doi.org/10.7860/JCDR/2016/ 18656.8283.

- Mohamed HG, Idris SB, Mustafa M, Ahmed MF, Åstrøm AN, Mustafa K, et al. Impact of Chronic Periodontitis on Levels of Glucoregulatory Biomarkers in Gingival Crevicular Fluid of Adults with and without Type 2 Diabetes. PLoS One. 2015;10(5):e0127660. https://doi.org/10.1371/journ al.pone.0127660.
- Mopidevi A, et al. Salivary visfatin concentrations in patients with chronic periodontitis: an analysis before and after periodontal therapy. Indian J Dent Res. 2019;30(6):864–9.
- Özcan E, Saygun NI, Serdar MA, Bengi VU, Kantarcı A. Non-Surgical Periodontal Therapy Reduces Saliva Adipokine and Matrix Metalloproteinase Levels in Periodontitis. J Periodontol. 2016;87(8):934–43. https://doi.org/ 10.1902/jop.2016.160046.
- Özcan E, Saygun NI, Serdar MA, Kubar A, Bengi VU. Porphyromonas gingivalis and Epstein-Barr Virus Are Associated With Increased Levels of Visfatin in Gingival Crevicular Fluid. J Periodontol. 2016;87(4):443–51.https:// doi.org/10.1902/jop.2015.150424.
- Özcan E, Saygun NI, Serdar MA, Kurt N. Evaluation of the salivary levels of visfatin, chemerin, and progranulin in periodontal inflammation. Clin Oral Investig. 2015;19(4):921–8. https://doi.org/10.1007/s00784-014-1308-0.
- Paul R, et al. Evaluation of association between Porphyromonas gingivalis and visfatin levels in chronic periodontitis patients. J Indian Soc Periodontol. 2020;24(6):530–4.
- 29. Pradeep AR, Raghavendra NM, Ramchandra Prasad MV, Kathariya R, Patel SP, Sharma A. Gingival crevicular fluid and serum visfatin concentration: their relationship in periodontal health and disease. J Periodontol. 2011;82(9):1314–9. https://doi.org/10.1902/jop.2011.100690.
- Pradeep AR, Raghavendra NM, Sharma A, Patel SP, Raju A, Kathariya R, et al. Association of serum and crevicular visfatin levels in periodontal health and disease with type 2 diabetes mellitus. J Periodontol. 2012;83(5):629–34. https://doi.org/10.1902/jop.2011.110272.
- Raghavendra NM, Pradeep AR, Kathariya R, Sharma A, Rao A, Naik SB. Effect of non surgical periodontal therapy on gingival crevicular fluid and serum visfatin concentration in periodontal health and disease. Dis Markers. 2012;32(6):383–8. https://doi.org/10.3233/DMA-2012-0897.
- Rezaei M, Bayani M, Tasorian B, Mahdian S. The comparison of visfatin levels of gingival crevicular fluid in systemic lupus erythematosus and chronic periodontitis patients with healthy subjects. Clin Rheumatol. 2019;38(11):3139–43. https://doi.org/10.1007/s10067-019-04708-w.
- Saljoughi F, Nasri K, Bayani M. Gingival crevicular fluid levels of visfatin in patients with chronic periodontitis and polycystic ovary syndrome. Obstet Gynecol Sci. 2020;63(1): 87–93. https://doi.org/10.5468/ ogs.2020.63.1.87.
- 34. Saseendran G, et al. Comparative evaluation of salivary visfatin levels in healthy and periodontally diseased patients before and after scaling and root planing. J Pharm Bioallied Sci. 2021;13(5):S624–8.
- Tabari ZA, Azadmehr A, Nohekhan A, Naddafpour N, Ghaedi FB. Salivary visfatin concentrations in patients with chronic periodontitis. J Periodontol. 2014;85(8):1081–5. https://doi.org/10.1902/jop.2013.130388.
- Tabari ZA, Ghaedi FB, Azadmehr A, Nohekhan A, Tabrizi MAA, Ardakani MRT, et al. Salivary Visfatin Concentration in Response to Non-surgical Periodontal Therapy. J Clin Diagn Res. 2015;9(4):ZC05–ZC08. https://doi. org/10.7860/JCDR/2015/11537.5773.
- Wu Y, Chen L, Wei B, Luo K, Yan F. Effect of non-surgical periodontal treatment on visfatin concentrations in serum and gingival crevicular fluid of patients with chronic periodontitis and type 2 diabetes mellitus. J Periodontol . 2015;86(6):795–800. https://doi.org/10.1902/jop.2015.140476.
- Ziaei N, Golmohammadi S, Ataee M, Ardalani F, Abbasi MM. Effect of non-surgical periodontal treatment on three salivary adipokines in diabetic patients with periodontitis. J Dent Res Dent Clin Dent Prospects. 2020;14(3):199–205. https://doi.org/10.34172/joddd.2020.029.
- Akram Z, et al. Cytokine Profile in Chronic Periodontitis Patients with and without Obesity: A Systematic Review and Meta-Analysis. Dis Markers. 2016;2016:4801418.
- 40. Akram Z, et al. Resistin as potential biomarker for chronic periodontitis: A systematic review and meta-analysis. Arch Oral Biol. 2017;73:311–20.
- Samal B, et al. Cloning and characterization of the cDNA encoding a novel human pre-B-cell colony-enhancing factor. Mol Cell Biol. 1994;14(2):1431–7.

- 42. Moschen AR, et al. Visfatin, an adipocytokine with proinflammatory and immunomodulating properties. J Immunol. 2007;178(3):1748–58.
- Luk T, Malam Z, Marshall JC. Pre-B cell colony-enhancing factor (PBEF)/visfatin: a novel mediator of innate immunity. J Leukoc Biol. 2008;83(4):804–16.
- Karampela I, Christodoulatos GS, Dalamaga M. The Role of Adipose Tissue and Adipokines in Sepsis: Inflammatory and Metabolic Considerations, and the Obesity Paradox. Curr Obes Rep. 2019;8(4):434–57.
- Saadoun MM, et al. Serum Visfatin as a Diagnostic Marker of Active Inflammatory Bowel Disease. J Gastrointestin Liver Dis. 2021;30(3):339–45.
- Polyakova YV, et al. Visfatin and Rheumatoid Arthritis: Pathogenetic Implications and Clinical Utility. Curr Rheumatol Rev. 2020;16(3):224–39.
- Radzicka S, et al. The role of visfatin in pathogenesis of gestational diabetes (GDM). Ginekol Pol. 2018;89(9):518–21.
- Dakroub, A., et al., Visfatin: A Possible Role in Cardiovasculo-Metabolic Disorders. Cells, 2020. 9(11).
- Kocher T, et al. Periodontal complications of hyperglycemia/diabetes mellitus: Epidemiologic complexity and clinical challenge. Periodontol 2000. 2018;78(1):59–97.
- Chen Z, et al. Research on the correlation of diabetes mellitus complicated with osteoporosis with lipid metabolism, adipokines and inflammatory factors and its regression analysis. Eur Rev Med Pharmacol Sci. 2017;21(17):3900–5.
- 51. Barros SP, et al. Gingival crevicular fluid as a source of biomarkers for periodontitis. Periodontol 2000. 2016;70(1):53–64.
- Buduneli N, Kinane DF. Host-derived diagnostic markers related to soft tissue destruction and bone degradation in periodontitis. J Clin Periodontol. 2011;38(Suppl 11):85–105.
- 53. Papagerakis P, et al. Saliva and Gingival Crevicular Fluid (GCF) Collection for Biomarker Screening. Methods Mol Biol. 2019;1922:549–62.
- 54. Cecoro, G., et al., Periodontitis, Low-Grade Inflammation and Systemic Health: A Scoping Review. Medicina (Kaunas), 2020. 56(6).

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